

# Computer simulation approaches in structural biology

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## Possible applications

- when the problem cannot be examined experimentally (transition states)
- to complement experimental data (eg. structure determinations)
- to improve resolution of experimental data (eg. build a model)
- to interpret experimental data (e.g. kinetic measurements on mutants)
- to study enzyme families (e.g. to study evolutionary relationships)
- **prediction** (structure, mutants, mechanism ...)

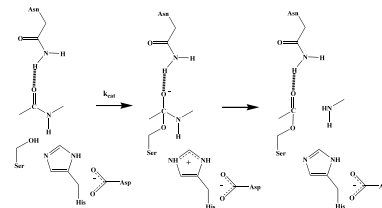
## Problems in structural biology

### 1. Enzymatic mechanisms

	enzyme	$k_{cat}$ [ $s^{-1}$ ]	water
carbonic anhydrase	$6 \times 10^5$	$10^{-9}$	
acetylcholine esterase	$2 \times 10^4$	$8 \times 10^{-10}$	
staphylococcal nuclease	$10^2$	$2 \times 10^{-14}$	

## 1. Enzymatic mechanisms

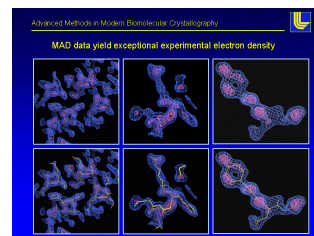
### Serine proteases



## Enzymatic mechanisms Problems

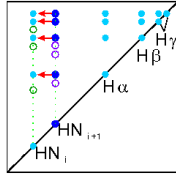
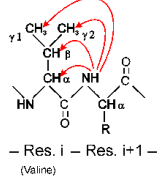
- Mechanism of action
  - description of the reaction steps
  - catalytic residues
  - rate limiting step
- Origin of catalytic power
  - proximity effect
  - steric stress
  - general acid/base catalysis
  - contribution of far-lying residues
  - gas phase like environment
  - Low Barrier Hydrogen Bonds
- Mutation effects

## 2. Structure determination X-ray crystallography



Structure determination  
NMR

Dipeptide Fragment



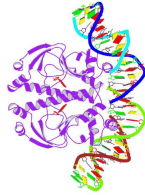
Structure determination

- bad parameter/data ratio  
model building  
refinement using force-fields  
multiple conformers (low energy?)
- identification of missing parts  
unstructured  
loops
- conformational changes  
substrate binding  
inhibitor binding  
cofactor binding

Examples  
DNA binding proteins

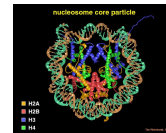
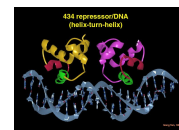
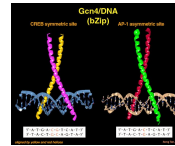


TATA box binding protein  
(TBP)



catabolic gene activator protein  
(CAP)

Examples  
DNA binding proteins



Quantitative applications

- dynamical analysis of protein (or complex) structures
- refinement of crystallographic or NMR data
- environmental effects (solution studies)
- kinetics of folding
- homology modelling (> 30%)
- vibrational spectra (low resolution)
- structural stability (differences)
- calculation of  $k_{cat}$  of enzymatic reactions
- $\Delta G$  of ligand binding (differences)
- mutational effects
- interpretation of catalytic effects

Validation

Comparison with experimental results

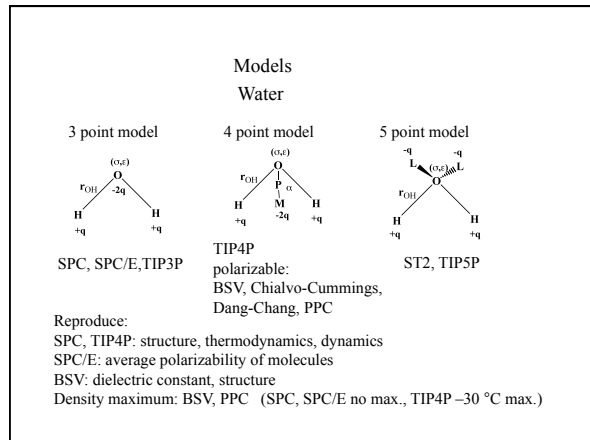
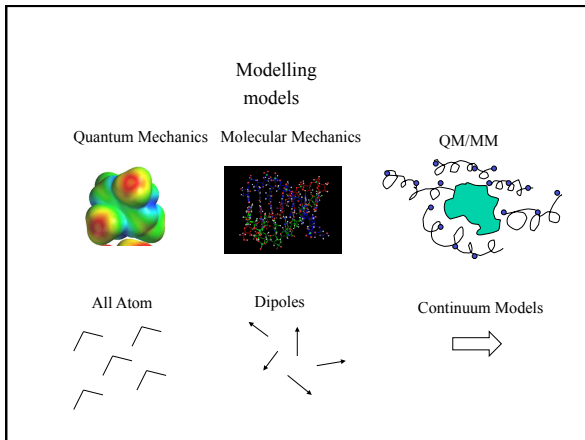
Structure determination

X-ray crystallography: comparison of computed and measured structure factors (R factor)

Comparison with homolog proteins  
likely mechanism of action

Interpretation of catalytic effects

reproducing  $k_{cat}$   
reproducing  $k_{cat}(\text{mutans})/k_{cat}(\text{nativ})$  data  
explaining pH dependence  
explaining dependence on ion strength  
prediction

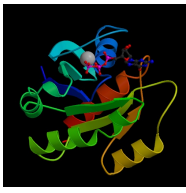


- ### Strategies
- Principles
- The model has to match with the chemical nature of the problem (QM is not always the best)
  - Consider the global problem. (define the questions)
  - All important parts have to be represented. (environmental effect)
  - profit/cost optima
  - **The error in different parts of the model has to be similar for the given problem.**

- ### Model building Errors
- wrong interpretation of the catalytic effect
  - wrong environment (e.g. missing ions)
  - small environment (e.g. *ab initio* calculations)
  - gas phase (neutrality)
  - incorrect water environment (crystallographic waters)
  - wrong treatment of electrostatic effects ( $\epsilon_{in}$ ,  $pK_a$ )
  - ignoring dynamic effects

### Model building Examples

Ras p21

$$GTP + H_2O \rightarrow GDP + HPO_4^{2-}$$


- What is the mechanism?
- Proton transfer pathways?
- Role of Lys-16?
- Role of Gln-61?
- Role of substrate in catalysis?
- Why cofactor is required?

- ### Ras p21 Possible mechanism
1. Activation of attacking nucleophile (water)?
- deprotonation by Gln-61
  - deprotonation by another water
  - not deprotonated (neutral water attacks)
  - concerted mechanism
  - $\gamma$ -phosphate is the general base
  - proximal Asp or Glu (Asp-57) deprotonates

### Ras p21 Mechanisms

#### Experimental results

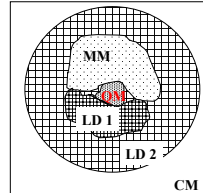
- Mutational data: Gln-61, Gly-12, Gly-13 drop in activity ( $10^{-3}$ )
- Structure: all are in proximity to  $\gamma$  phosphate
- NMR: pKa of the  $\gamma$  phosphate is affected by the mutations
- Kinetics:  $k_{cat}$  const. if pH > 3, and dependent on pKa of the  $\gamma$  phosphate



Hypothesis: catalysis occurs with substrate assistance

### Ras p21 Studies on mechanism

Warshel et al. Nat. Struct. Biol. (1994) 1 pp. 476 – 484.



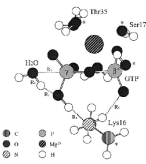
Method: EVB/FEP

Activation barrier calculation for all possible mechanisms including all sidechains and the whole environment

Conclusion: it is the  $\gamma$  phosphate that deprotonates the attacking water (E)

### Ras p21 Studies on mechanism

Futatsugi et al. *Biophys. J.* (1999) 77 3287-3292.



#### Method:

- structure: optimized enzyme
- *ab initio* calculations on ground and transition state

#### Conclusion:

- "proton ping-pong" with the assistance of Lys
- Gln helps to position of the catalytic water

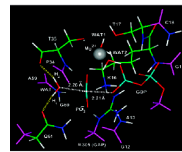
#### Error:

- assume the protonation state of Lys and GTP
- missing enzymatic environment

### Ras p21 Studies on mechanism

Cavalli et al. *J. Am.Chem.Soc.* (2002) 124, pp.3763-3768.

DFT-MD (Carr-Parinello)



- 85% of the electric field of the protein is represented
- 2 ps MD 60000 CPU hours
- $O_{\gamma}$ -Py 1.8 Å distant constraints
- when released proton jumps to O of Gln61

#### Error:

irreal pKa values,  
no comparison with solution reaction  
Gln→Glu mutation cannot be explained

- $AlF_3$  TS analog